Comparison of Two Methods for the Measurement of Methyl Mercury Concentrations in Penobscot River Sediments

Dr. R.J. Flett, *Flett Research Ltd*; Ms. Brenda Lasorsa, *Battelle Marine Sciences Laboratory*; Dr. Gary Gill, Battelle Marine Sciences Laboratory; Dr. Holger Hintelmann, Trent University

Poster presented at the 10th International Conference on Mercury as a Global Pollutant (ICMGP), Halifax, Nova Scotia, Canada, July 24-29, 2011

Introduction

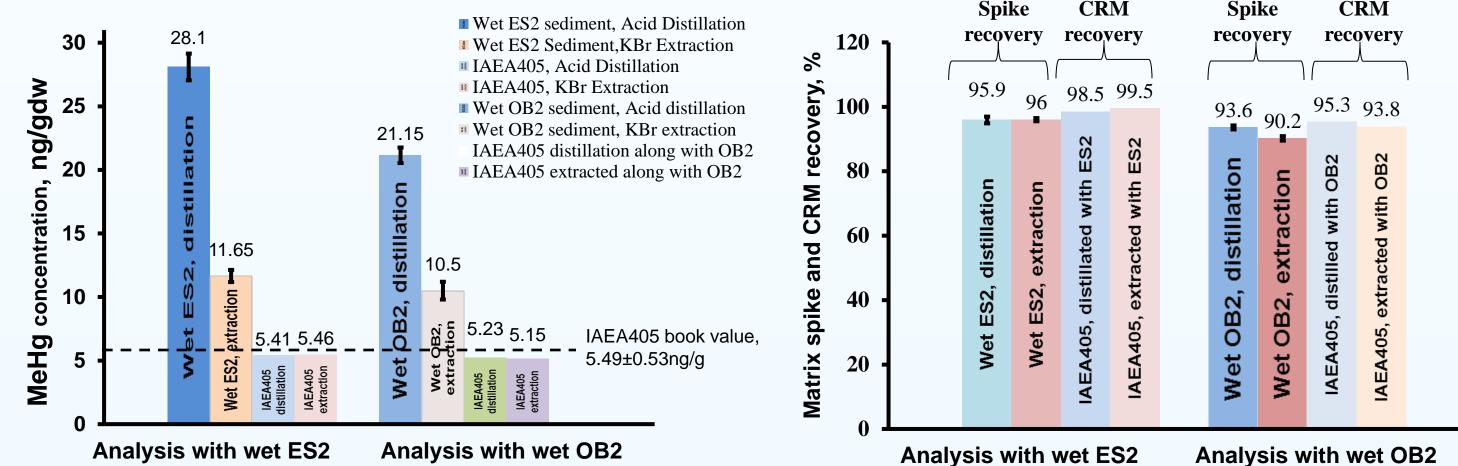
Acid distillation and solvent extraction methods^{1,2,3,4} are widely used to determine monomethyl mercury (MMHg) content in sediments. During initial phases of the Penobscot River Mercury Study in Maine, USA in 2006 - 07, both methods were evaluated for performance. A combination of results from 3 laboratories (Flett Research Ltd., Battelle Marine Sciences Laboratory and Trent University) indicated the following:

- Results from solvent extraction average $\sim 50\%$ lower than acid distillation (number of tested samples >200);
- CRM IAEA-405 and matrix spike recoveries were similar and good with both methods;
- Artifact generation from conversion of Hg^{2+} to MMHg in acid distillation^{2,4,5} is low (<0.1%) according to isotopic labeling method⁶ and non-isotopic standard addition Hg²⁺ spike recovery and therefore is not a major factor for higher distillation results.

The cause of the discrepancy was unclear and therefore it was decided to test the following hypothesis: **Distillation** is providing the correct methyl mercury concentrations and the application of heat and/or acid is required to release methyl mercury for accurate determination by the solvent extraction method.

Results (Continued)

3. Solvent extraction MMHg results on ES2 and OB2 wet sediment are about half of those determined by distillation even though Matrix Spike recoveries and CRM IAEA405 recoveries were all better than 90% with either method.

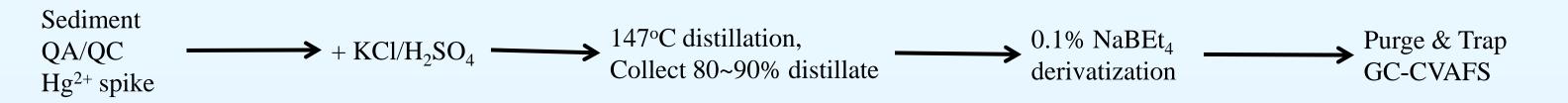


Two large well-homogenized sediment samples (one river and one estuary, taken as part of QA/QC procedures for the Penobscot Study) were used to test this hypothesis.

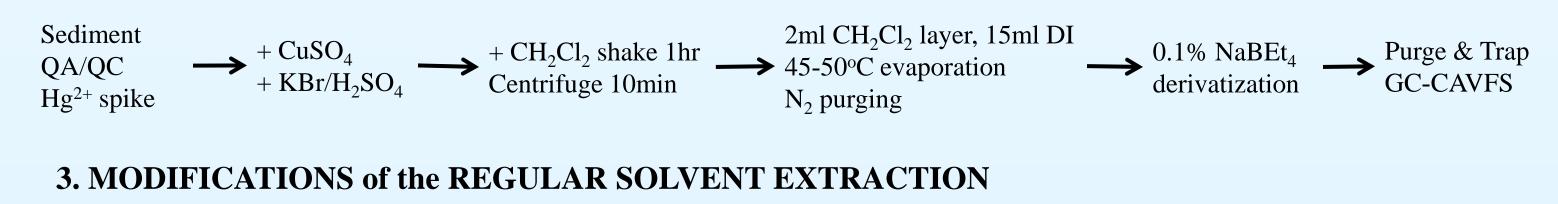
Methods

Sediments from two sites in Penobscot River, Maine, USA were sampled in this study. ES2 was obtained from the river estuary, whereas OB2 was taken upstream from the estuary. In the flow chart below, DI is ion-exchanged pure water, KCl/H₂SO₄ is 1.6% KCl in 8 M H₂SO₄ solution, CuSO₄ is 1M CuSO₄ solution, KBr/H₂SO₄ is 18% KBr in 10% H₂SO₄ solution, CVAFS is cold vapor atomic fluorescence spectrometry, QA/QC includes MMHg matrix spike, inorganic Hg²⁺ spike and sediment CRM IAEA405. When wet sediments were analysed, all MMHg concentration was normalized into dry weight basis, as ng/gdw.

1. DISTILLATION with KCl/H₂SO₄



2. REGULAR SOLVENT EXTRACTION with KBr/H₂SO₄/CH₂Cl₂



Analysis with wet OB2

4. Sonication pretreatment of wet ES2 sample before going through solvent extraction didn't improve MMHg recovery. The MMHg detected was still half of that determined by distillation.

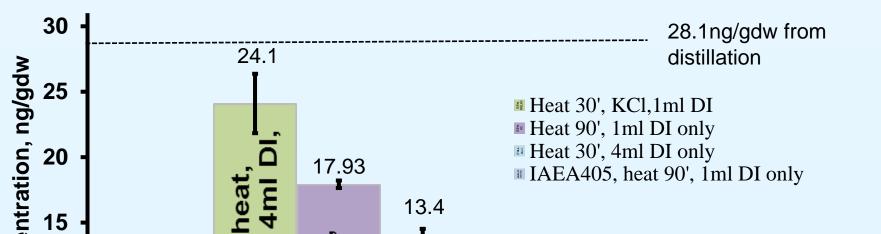
5. After a solvent extraction of wet ES2, the previously extracted sediment was distilled or went through a 2nd solvent extraction.

Sample Type	MMHg detected in Sediment by Distillation, ng/gdw	MMHg detected in sediment by KBr extraction, ng/gdw	MMHg detected in previously KBr solvent extracted sediment (converted to equivalent MMHg concentration in dry sediment)	
			Distillation, ng/gdw	KBr Extraction, ng/gdw
Wet ES2	28.1	12.4	7.61	2.23
Wet ES2 with MMHg spike		42.55	7.35	4.13
IAEA405 (5.49±0.53 ng/g)	5.41	4.48	0.31	0.40
Hg ²⁺ conversion to MMHg	0.07%	0.03%	0.08~0.16%	<0.01%

The distillation of previously extracted sediment detected 7.61 ng/gdw MMHg, indicating a significant amount of MMHg remained after extraction. On the other hand, the distillation of previously extracted sediment which contained a matrix spike found a similar amount of MMHg (7.35) as in the unspiked sample. This indicated the aqueous based MMHg in matrix spike is easily extracted into CH₂Cl₂ (>90% recovery was found) whereas ambient MMHg in ES2 is poorly extracted.

6. Heat pretreatment of sediments at 100°C prior to solvent extraction

6.1 Heat ES2 wet sediment with KCl/H₂SO₄ in DI only OR



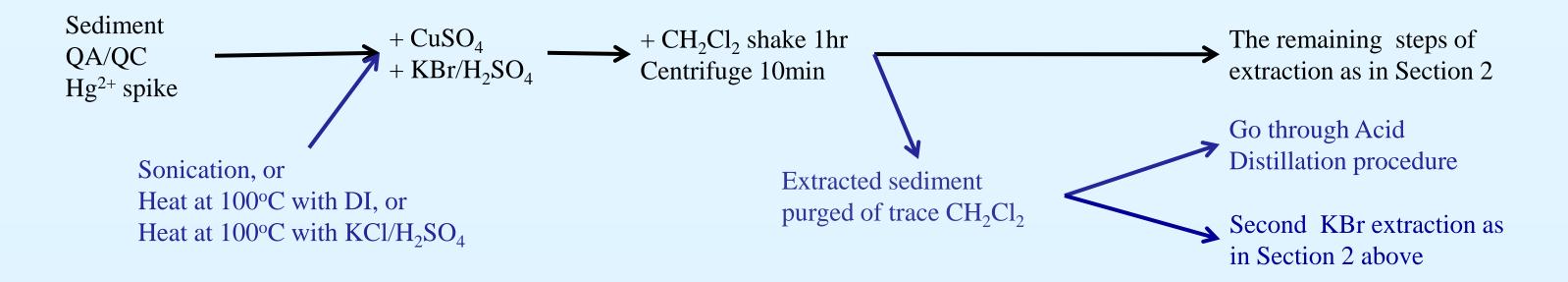
• The matrix spike and CRM recoveries were good.

• MMHg solvent extraction recovery was highest when sediment was pretreated

3.1 Distill the previously extracted sediment

3.2 Extract the previously extracted sediment

3.3 Sample pretreatment with 100°C heat and DI OR 100°C heat and KCl/H₂SO₄ OR Sonication

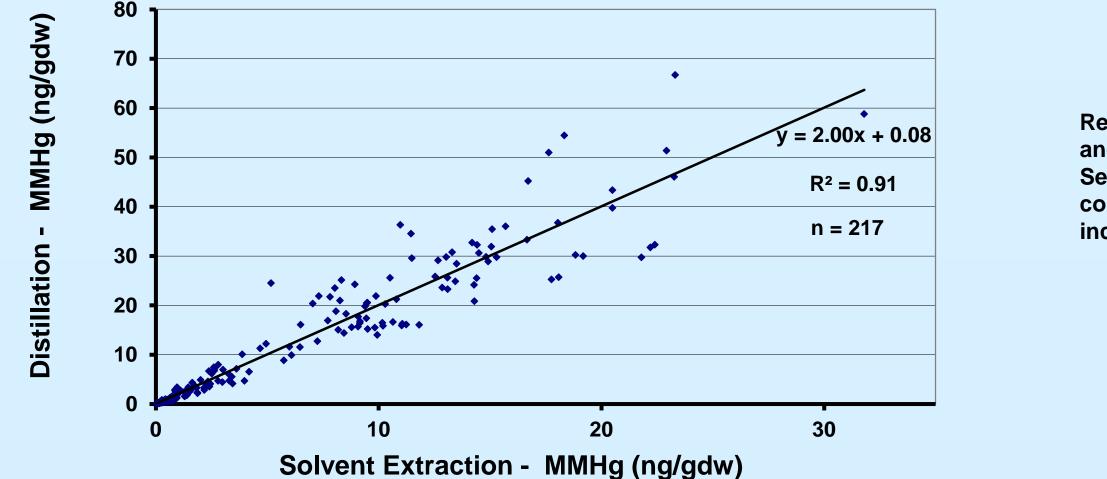


4. INORGANIC Hg²⁺ SPIKE CONVERSION TO MMHg

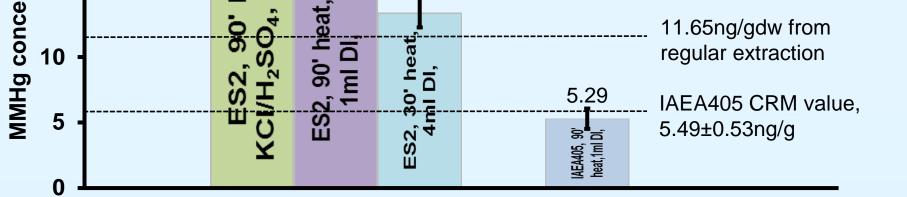
For each experiment above, External Hg²⁺ spike was included to evaluate the MMHg conversion. Hg²⁺ spike is more than 1000 X MMHg concentration measured from Acid Distillation.

Results

1. Some of the Penobscot River Mercury Study data that prompted the present investigation. All results corrected for spike recovery. Acid distillation recovered MMHg at an average of <u>2.13 times</u> of that recovered by solvent extraction.



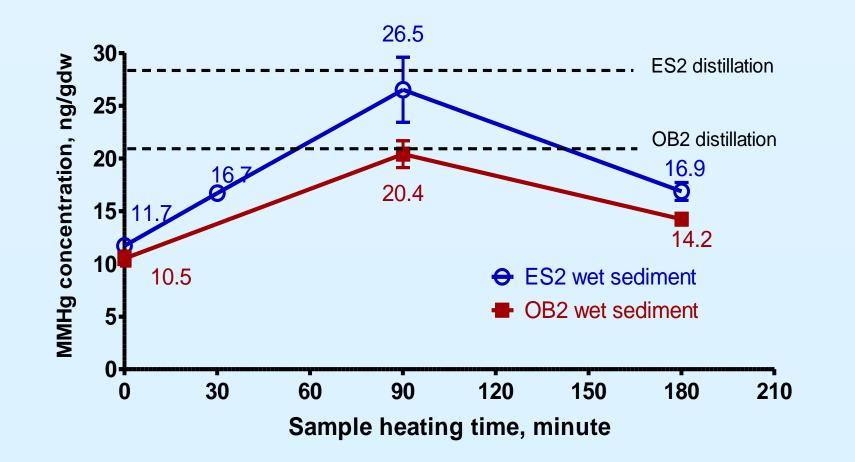




Solvent extraction with different sample pretreatment

~0.4g ES2 wet sediment, heat with 0.2ml KCl/H₂SO₄ in 4ml DI or without KCl in 1 or 4 ml DI before extraction.

6.2 Effect of different heating times on ES2 and OB2 wet sediment with KCl/H₂SO₄



Wet ES2 and OB2 showed similar trends of heating effect.

Comparative results from Distillation are 28.1ng/gdw and 21.2 ng/gdw for ES2 and OB2 wet, respectively.

~0.4g ES2 and OB2 wet sediment, heat with 0.2ml KCl/H₂SO₄ in 1ml DI (not 4ml) before extraction.

Conclusions

• MMHg determined by KBr Solvent Extraction underestimates the real MMHg concentration in Penobscot River sediment samples by ~50% due to low extraction efficiency. A portion of MMHg in these sediments is in a form that is different than the MMHg in standard spike solutions, making it unavailable to solvent extraction at room temperature: matrix spike recovery is good but final measurement is low. Good spike recovery does not guarantee good sample recovery. • Pretreatment of sediment with heat, and particularly heat in the presence of KCl/H₂SO₄, significantly improves the solvent extraction process. Although the conversion of inorganic Hg²⁺ is correspondingly increased, from 0.03% to 0.13%, the artifact accounts for less than 7% of the overall MMHg detected, and is only significant when THg/MeHg ratio is extremely high.

with heat @ 100°C with KCl/H₂SO₄.

• If preheated in DI only, MMHg detected were higher than regular KBr/CH₂Cl₂ extraction, but still significantly lower than that from distillation.

• The MMHg conversion from Hg²⁺ with this heating process was 0.07%.

2. Trace level isotopic Hg labeling experiments showed 0.01% to 0.04% inorganic Hg²⁺ converted into MMHg while high level non-isotopic Hg²⁺ spike experiments showed 0.07% conversion occurred in the distillation of Penobscot River sediments, vs 0.03% found in extraction. The artifact MMHg determined by isotopic labeling procedure was less than 2.4% of the MMHg value determined by the distillation method.

Sample Station	Sample Description	Ambient THg*, ng/gdw	Ambient MMHg, ng/gdw	Average ²⁰⁰ Hg methylated (artifact), %
OV5 (n=3)	freshwater	51	0.72	0.01
OB2 (n=3)	tidal river	1022	25.0	0.02
OB4 (n=6)	tidal river	1078	15.1	0.03
ES2 (n=6)	estuary	1622	27.3	0.04

*: THg results listed here are site average of 2007 from all labs

ES2 and OB2 sediment samples used in this study have Total Hg of 892 and 1185 ng/gdw respectively, MMHg (by distillation) of 28.1 and 21.2 ng/gdw respectively. With typical 0.07% conversion found in distillation, the artifact MMHg generated (as a % of ambient) is 2.2% for ES2 and 3.9% for OB2. This artifact is not a major contributor to the large discrepancy between the two methods.

• Poor solvent extraction is matrix specific. Solvent extraction of CRM IAEA-405 is nearly complete while test sediments ES2 and OB2 extractions were only about 50%. Good CRM recovery does not guarantee good sample recovery.



The Penobscot River Mercury Study is thanked for permitting these data to be presented here.



1. M. Horvat, Water Air Soil Pollut. 56 (1991) 95; 2. M. Horvat, N.S. Bloom, L. Liang, Anal. Chim. Acta 281 (1993) 135; 3. M. Horvat, N.S. Bloom, L. Liang, Anal. Chim. Acta 282 (1993) 153; 4. Bloom, N.S., Coleman, J.A., and Barber, L., Fres. Anal. Chem. 358 (1997) 371; 5. Karl C. Bowles, Simon C. Apte, Anal. Chim. Acta 419 (2000) 145; 6. H. Hintelmann, Chemosphere 39 (1999) 1093